Contents lists available at ScienceDirect



Review

Pharmacological Research



journal homepage: www.elsevier.com/locate/yphrs

Dioscin: A diverse acting natural compound with therapeutic potential in metabolic diseases, cancer, inflammation and infections



Xufeng Tao, Lianhong Yin, Lina Xu, Jinyong Peng*

College of Pharmacy, Dalian Medical University, Western 9 Lvshunnan Road, Dalian, 116044, China

| ARTICLE INFO | ABSTRACT | | | | |
|--------------------|--|--|--|--|--|
| Keywords: | Currently, the numbers of patients with cancer, fibrosis, diabetes, chronic kidney disease, stroke and osteo- | | | | |
| Dioscin | porosis are increasing fast and fast. It's critical necessary to discovery lead compounds for new drug develop- | | | | |
| Metabolic disorder | ment. Dioscin, one active compound in some medicinal plants, has anti-inflammation, immunoregulation, hy- | | | | |
| Cancer | polipidemic, anti-viral, anti-fungal and anti-allergic effects. In recent years, dioscin has reached more and more | | | | |
| Diabetes | attention with its potent effects to treat liver, kidney, brain, stomach and intestine damages, and metabolic | | | | |
| Oberity | diseases including diabetes, osteoporosis, obesity, hyperuricemia as well as its anti-cancer activities through | | | | |
| Obesity | adjusting multiple targets and multiple signals. Therefore, dioscin is a promising multi-target candidate to treat | | | | |
| | various diseases. This review paper summarized the progress on pharmacological activities and mechanisms of | | | | |

1. Introduction

In recent years, the numbers of patients with cancers, fibrosis, liver injury, cirrhosis, diabetes, chronic kidney disease, and osteoporosis are increasing (Table 1) [1]. Thus, development and exploration of new treatment modalities to reduce the burden and control the diseases are critical important.

Evidences have shown that drugs or functional products with single target have less treatment actions and more side effects when they are used to treat complex diseases involving multiple pathological changes [2,3]. In response to these challenges, there are growing interests to develop multi-target drugs or functional products. Traditional Chinese medicines (TCMs) have been used to treat various diseases in China for thousands of years [4,5]. In the past decades, the works of TCMs on multiple-target effects have significantly increased. Beyond that, TCMs process some positive features including diversity, broad continuity, relatively low cost, low side effects and high efficiency [6]. A large number of pharmacological researches on active natural products derived from TCMs have achieved tremendous progress in recent years (Fig. 1A). The works of artemisinin against malaria by Youyou Tu have won the "2015 Nobel Prize in Hysiology or Medicine". Therefore, it's feasible to discovery natural products from TCMs for development and exploration of innovative drugs.

Dioscin, a steroidal saponin, is an active ingredient in some TCMs including Liuwei Dihuang decoction and Di'ao Xinxue kang [7-10]. The

data from "PubMed" and "CNKI" databases have indicated that the works on dioscin have markedly increased in recent years (Fig. 1B). As shown in Fig. 2, dioscin also is one active compound in Dioscoreae rhizoma (Shanyao in Chinese, one famous vegetable), and several Dioscoreaceae plants including Dioscorea Zingiberensis C.H. Wright and Dioscorea nipponica Makino [11,12]. The book of "Sheng Nong's Herbal Classic" has recorded that "Dioscorea is sweet to level the smell with the characteristics of reducing visceral injury and reinforcing insufficiency", which can also remove cold- and heat-evil, replenish Qi and promote muscle growth. In addition, Dioscorea can increase the sensitivity of hearing and vision, and extend life expectancy [13]. Pharmacological tests have shown that dioscin has anti-tumor, antiinflammation, immuneregulation, hypolipidemic, anti-viral, anti-fungal and anti-allergic effects. Furthermore, some works in recent years have suggested that dioscin can protect liver, kidney, brain and gastrointestinal damages, and regulate the metabolic diseases including diabetes, osteoporosis, obesity and hyperuricemia through adjusting multiple targets and signals. Therefore, dioscin is a promising multi-target candidate to treat some diseases, suggesting that this natural product should have expansive application prospects.

dioscin, which may provide useful data for development and exploration of this natural product in the further.

Although these works have confirmed the pharmacological actions of dioscin, most of them focus on the effects in one disease or one mechanism. Therefore, we are trying to give a timely and comprehensive update on the pharmacological actions and targets of dioscin. References on this natural product have implemented by collecting

* Corresponding author.

E-mail address: jinyongpeng2005@163.com (J. Peng).

https://doi.org/10.1016/j.phrs.2018.09.022

Received 12 July 2018; Received in revised form 19 September 2018; Accepted 20 September 2018 Available online 10 October 2018

1043-6618/ © 2018 Elsevier Ltd. All rights reserved.

Table 1

Deaths, YLDs, and DALYs attributable to various diseases (1990-2013).

| Cause | Death rank | Deaths per 100,000 | YLD rank | YLDs per 100,000 | DALY rank | DALYs per 100,000 |
|---|------------|--------------------|----------|------------------|-----------|-------------------|
| Cardiovascular diseases | 1–1 | 231.3-241.4 | 11–9 | 211.2-295.6 | 3–1 | 4872.5-4601.7 |
| Neoplasms | 3–2 | 106.6-114.9 | 16–16 | 69.8–94.4 | 4–3 | 2873.4-2750.8 |
| Chronic respiratory diseases | 4–4 | 65.7–59.6 | 7–7 | 468.7-539.0 | 8-10 | 1900.0-1573.1 |
| Diabetes, urogenital, blood, and endocrine diseases | 8–5 | 29.6-41.2 | 5–4 | 714.7-915.0 | 12-8 | 1660.0-1976.6 |
| Digestive diseases | 11-13 | 19.8-16.3 | 14-14 | 128.2-118.0 | 18–17 | 747.4-521.4 |
| Neurological disorders | 13–9 | 19.2-27.6 | 6–5 | 700.3-828.5 | 15-13 | 1021.2-1173.0 |
| Cirrhosis | 14-12 | 15.8-17.0 | 21-21 | 7.9–7.6 | 19–18 | 511.5-514.4 |
| Other non-communicable diseases | 15–15 | 15.7-10.4 | 3–3 | 1535.0-1646.2 | 17–19 | 753.6-378.4 |
| Mental and substance use disorders | 19–19 | 3.5-3.9 | 1–1 | 2101.7-2258.4 | 7–5 | 2248.5-2417.0 |
| Musculoskeletal disorders | 21-20 | 1.2-1.6 | 2-2 | 1713.0-2040.9 | 11–7 | 1753.7-2085.7 |
| Diarrhea, lower respiratory, and other common infectious diseases | 2–3 | 148.4-66.3 | 10-11 | 268.8-200.0 | 1-2 | 10356.1-3487.2 |
| Neonatal disorders | 5–7 | 64.7-28.6 | 15-12 | 103.0-176.5 | 2–4 | 5684.9-2646.2 |
| HIV/AIDS and tuberculosis | 6–6 | 39.0–36.7 | 17–15 | 45.1–76.7 | 13–9 | 1612.8-1663.4 |
| Neglected tropical diseases and malaria | 10-14 | 20.6-13.9 | 8–10 | 390.9-280.5 | 9–12 | 1848.6-1265.6 |
| Nutritional deficiencies | 16–16 | 14.2-9.5 | 4–6 | 868.2-582.9 | 10-15 | 1777.7-1044.5 |
| Other communicable, maternal, neonatal, and Nutritional diseases | 17–17 | 10.1-5.7 | 18-18 | 61.0-49.8 | 5–6 | 2766.2-2385.9 |
| Maternal disorders | 18-18 | 7.1-4.1 | 19–19 | 21.4-18.7 | 20-20 | 428.3-251.6 |
| Unintentional injuries | 7–8 | 38.0-28.0 | 9–8 | 345.0-302.2 | 6–11 | 2510.7-1478.6 |
| Transport injuries | 9–10 | 21.7-20.7 | 12-13 | 180.3-142.3 | 14-14 | 1338.5-1101.5 |
| Self-harm and interpersonal violence | 12-11 | 19.8-17.4 | 20-20 | 19.4–14.7 | 16–16 | 971.2-789.6 |
| Forces of nature, war, and legal intervention | 20-21 | 1.99–0.70 | 13–17 | 151.5–54.9 | 21–21 | 257.1-85.3 |

YLDs = years living with disability. DALYs = disability-adjusted life-years.

Communicable and neonatal diseases (column 1-10), Non-communicable diseases (column 11-17), Injuries (column 18-21).



Fig. 1. The papers about (A) TCMs and (B) dioscin researches published in "PubMed" and "CNKI" databases from 1997 to 2017. The data from "PubMed" and "CNKI" databases have indicated that the works on traditional Chinese medicines (TCMs) and dioscin have achieved tremendous progress in recent years.

English literatures (PubMed, Elsevier ScienceDirect, Web of Science and Medline databases). The purpose of the review paper was to investigate and summarize the latest advances of pharmacological activities and functional mechanisms of dioscin, and then provide useful data for development and exploration of this natural product in the further.

2. Mechanisms and anti-tumor effects of dioscin

At present, cancer caused by a lot of factors is expected to exceed cardiovascular diseases to be the most important cause of mortality in next few years [14,15]. Although some chemical drugs are effective to treat cancer, a large number of serious side effects have found. Therefore, development of new candidates with high efficiency and low side effects is urgent.

2.1. Anti-breast cancer effect and mechanism of dioscin

Breast cancer is the most common cancer in worldwide with an estimated 1.7 million cases and 521,900 deaths in 2012. The survival rates of the breast cancer are currently estimated to be less than 5-10% for 10-year and 25% for 5-year [16]. In breast cancer, GATA-binding protein 3 (GATA3) has the ability to reverse epithelial mesenchymal transition in invasive breast cancer cells, which can result in inhibiting metastasis. Aumsuwan, P. et al., have found that dioscin can inhibit cellular invasion in human MDA-MB-231 breast cancer cells by upregulating the expression of GATA3 [17]. In addition, microarray asays have shown that dioscin can up-regulate 36 genes and down-regulate 60 genes in MDA-MB-231 cells, as well as up-regulate 395 genes and down-regulate 406 genes in MCF-7 cells [18]. These data suggest the multiple networks and pathways associated with the anti-cancer effects of dioscin. After the appearance of apoptosis in breast cancer cells, apoptosis-inducing factor (AIF) translocates to the nucleus and participates in caspase-independent apoptotic events including chromatin condensation and large-scale DNA fragment. Dioscin can induce the death of MDA-MB-453, MDA-MB-231 and T47D cells via activating AIFfacilitating caspase-independent pathway and reducing the espression levels of anti-apoptotic related proteins [19]. In clinical, resistance to chemotherapeutic agents, especially to multi-drug resistance (MDR), is a major cause of treatment failure [20]. Importantly, regulating MDR1 level can up-regulate p-glycoprotein (p-gp) expression and suppress autophagy in cancer cells [21,22]. Dioscin can increase adriamycin (ADR) chemosen- sitivity by decreasing MDR1 levels, and improve the cytotoxicity of ADR through inducing autophagy [23].

2.2. Anti-gastric carcinoma effect and mechanism of dioscin

Owing to the high mortality and low survival rate, gastric cancer has become the second leading cause of cancer-related deaths [24]. Hu, M.M. et al., have investigated that dioscin processes potential actions against human gastric cancer through inducing cell apoptosis via activating the extrinsic and mitrochondrial pathways [25]. Loss of cell



Fig. 2. Dioscin presents in several medicinal plants including Dioscoreae rhizoma, Dioscorea nipponica Makino, Dioscorea hypoglauca Palibin, Dioscorea bulbifera L, Dioscorea Zingiberensis C.H. Wright and Dioscorea hispida. The images of these medicinal plants are obtained from the internet.

cycle control can lead to the proliferation of uncontrolled cells for the development of cancer. Therefore, the selective cyclin dependent kinase (CDK) inhibitor is one potent agent to treat cancer [26]. Dioscin can induce G2/M phase arrest and apoptosis in SGC-7901 cells by inhibiting CDK-activating kinase activity [27]. Moreover, Zhao, X.W. et al., have found that dioscin shows cytotoxicity to human MGC-803 gastric cancer cells and significantly inhibits tumor growth of xenografts in nude mice via including DNA damage and cell cycle arrest [28]. In addition, isobaric tags for relative and absolute quantification (iTRAQ)-based proteomics approach has found 121 differentially expressed proteins associated with cell apoptosis, cycle and migration caused by dioscin. Recent studies have also indicated that long non-coding RNAs (lncRNAs) are involved in the development of cancer [29]. In the study of Ma, T. et al., high expression level of lncRNA HOTAIR has been found in gastric cancer tissue, which can be significantly down-regulated by dioscin [30].

2.3. Anti-liver cancer effect and mechanism of dioscin

Approximately 790,000 new cases and 818,000 deaths of liver cancer worldwide in 2013 have been reported [31]. Currently, the therapeutic methods including liver resection, palliative intra-arterial therapies and transplantation have been used. However, long-term prognosis of liver cancer remains poor because of chemotherapy -resistant and high tumor recurrence [32]. Dioscin can dose-dependently enhance caspase-3-dependent cell apoptosis in Huh7 cells [33]. The works of Zhang, G.X. et al., have also demonstrated that dioscin can inhibit the proliferation of hepatocellular carcinoma cells via inducing morphological changes and DNA damage in Bel-7402 cells [34]. No-tably, dioscin can also suppress tumor growth via inducing apoptosis in nude mice bearing Bel-7402 cells. In addition, dioscin can effectively reverse multidrug resistance in HepG2/ADR cells via inhibiting the activity of MDR1 promoter and P-gp function [35].

2.4. Anti-acute myeloid leukemia effect and mechanism of dioscin

Acute myeloid leukemia is the most common form of adult leukemia, and little progression has been made on developing effective therapies. Mitogen-activated protein kinases (MAPKs) exert vital biological effects on apoptotic signals in myeloid leukemia [36]. In the study of Wang, Y. et al., dioscin can notably induce the apoptosis of myeloblast leukemia HL-60 cells through activating c-Jun N-terminal kinase (JNK) and p38 MAPK [37]. In addition, the action of dioscin on ADR-resistant erythroleukemic cells (K562/ADR) has been tested by Wang, L.J. et al., and dioscin can reverse ADR-induced MDR through reducing the mRNA and protein levels of MDR1 and NF- κ B [38].

2.5. Anti-lung cancer effect and mechanism of dioscin

Lung cancer with high incidence, rapid progression and poor prognosis is the leading cause of cancer-related death. Wei, Y.L. et al., have proved that dioscin can significantly induce cell apoptosis on human A549, NCI-H446 and NCI-H460 lung cancer cells [39]. Furthermore, Hsieh, M. J. et al., have found that autophagy induced by disocin maybe one pathway for cell survival against apoptosis [40]. In addition, epithelial-to-mesenchymal transition (EMT) is a key cellular process during cancer development. Dioscin can suppress the migration and invasion of A549 lung cancer cells by inhibiting transforming growth factor- β 1 (TGF- β 1)-induced EMT [41]. Moreover, tyrosine kinase inhibitors (TKIs) exhibit good clinical benefits in the treatment of lung adenocarcinoma. The works of Wang, Y.C. et al., have indicated that dioscin can act as a dual inhibitor of MEK/ERK and PI3K/AKT signaling pathways to overcome TKIs resistance [42].

2.6. Anti-renal cancer effect and mechanism of dioscin

Total of 63,990 new cases and 14,400 deaths of renal cancer in 2017 have been found. TNF-related apoptosis inducing ligand (TRAIL) can

induce the apoptosis of tumor cells. The study of Kim, Y.S. et al., have found that dioscin can significantly enhance TRAIL-induced apoptosis [43]. Gap junction plays key roles in growth, differentiation and apoptosis on cancer cells. Zhang, G.Y. et al.,have indicated that dioscin can promote the function of gap junction in human renal carcinoma cell [44].

2.7. Anti-melanoma tumor effect and mechanism of dioscin

Melanoma has the features of high mortality, high invasion and high metastasis. Three major pigment enzymes including tyrosinase, tyrosinase-related protein (TRP)-1 and TRP-2 play important roles in melanoma tumor [45], and their activites can be regulated by microphthalmia-related transcription factor (MITF) [46]. Dioscin can inhibit α -melanocyte-stimulating hormone (α -MSH)-induced melanogenesis in B16 cells via decreasing the expression levels of tyrosinase, TRP-1, TRP-2 and MITF [47]. Connexin 43 (Cx43) is an important gap junction protein in tumor micro- environment (TME), and the works of Kou, et al., have found that dioscin can target Cx43 to activate macrophage sensitivity and inhibit tumor cell malignancy [48].

2.8. Anti-prostatic cancer effect and mechanism of diosicn

The patients with prostate cancer usually accept surgery or antiandrogen therapy at the early stage [49]. Prostate cancer stem cells (PCSCs) are the cell origin of prostatic cancer, which paly important roles in tumorigenesis. Our works have confirmed that dioscin has potent inhibitory activity on PC3 cell-derived mammos- pheres via activating estrogen receptor- β (ER β) [50].

2.9. Other anti-cancer effects and mechanism of dioscin

Epithelial ovarian cancer is the leading cause of gynecologic cancerassociated deaths among women, and cell apoptosis is closely related to the occurrence, progress and metastasis of the cancer. Dioscin can doseand time-dependently inhibit cell proliferation and induce cell apoptosis in human ovarian cancer SKOV3 cells [51]. ROS accumulation can cause cell apoptosis, mitochondrial damage, cytochrome C release and programmed cell death-5 (PDCD5) nuclear translation. Wang, Z.Y. et al., have found dioscin can induce ROS generation and apoptosis in human Kyse510 esophageal cancer cells through increasing oxidative stress via downregulating the espression levels of peroxiredoxin (PRDX)-1 and PRDX-6 [52]. Laryngeal cancer is the common malignant tumor of head and neck. In the works of Si, L.L. et al., dioscin causes cell apoptosis and DNA damage, up-regulates ROS level, induces S-phase arrest, and reduces invasion in human HEp-2 and TU212 laryngeal cancer cells [53]. The actions of dioscin on human cervical carcinoma HeLa and SiHa cells have been detected by Zhao, X.W. et al., and dioscin can cause cell apoptosis through regualting ROS-mediated DNA damage and mitochondrial signaling pathway [54]. Glioblastoma multiforme is a common malignant brain cancer, and dioscin exerts a promising inhibiting effect on glioblastoma cells via ehancing ROS accumulation, DNA damage and mitochondrial signals [55]. In addition, dioscin can markedly suppress tumor size and increase survival rate in glioma in vivo rat allograft model. Pancreatic cancer is one of the most lethal solid malignancies in the Western world. Recently, microRNA microarray analysis has been used to determine the underlying mechanisms of dioscin against pancreatic cancer, and total of 107 micro-RNAs with differential changes have been found to prove that dioscin exerts excellent activity against pancreatic cancer via miR-149-3 P/ Akt1 signaling pathway [56]. In recent years, the incidents of bladder carcinoma in China have increased rapidly. Dioscin can induce demethylation of death-associated protein kinase-1 (DAPK-1) and ras-association domain family 1 isoform A (RASSF-1) to cause apoptosis on T24 cancer cells [57]. Song, X.L. et al., have checked the actions of dioscin on NOZ and SGC996 gallbladder cancer cells, and dioscin can significantly inhibit cell proliferation and migration [58]. Colorectal cancer is the second leading cause of cancer-related deaths in the USA. Li, S. et al., have found that dioscin can induce the apoptosis of colon cancer cells through adjusting the phosphorylation levels of JNK and p38-MAPK [59]. Osteosarcoma is one common primary bone tumor. Liu, W.H. et al., have adopted in vitro and in vivo models, and dioscin inhibits osteosarcoma stem- cell-like properties and tumor growth via repressing Akt/glycogen synthase kinase 3 (GSK3)/ β -catenin pathway [60].

Together, the effects of dioscin against breast cancer, gastric carcinoma, acute myeloid leukemia, lung cancer, ovarian cancer, renal cancer, melanoma tumor, esophageal cancer, prostatic cancer, laryngeal cancer, cervical carcinoma, liver cancer, glioblastoma, pancreatic cancer, gallbladder cancer and bladder cancer have attracted more attentions, and these studies have described the mechanistic actions of dioscin on apoptosis, ROS generation, Ca^{2+} release, cell cycle and DNA damage.

3. Protective effects and mechanisms of dioscin against organ damages

3.1. Hepatoprotective effects of diosicn

Liver diseases are the major global public health problems in the world [61]. Among them, acute liver injury caused by adverse drug reactions accounts for more than 50% of the cases [62]. Moreover, liver can be damaged by many toxic chemicals including carbon tetra-chloride (CCl₄), alpha-naphthylisothiocyanate (ANIT) and dimethylnitrosamine (DMN) [63]. In addition, chronic liver diseases induced by viral infection, alcohol abuse, and obesity can cause hepatic encephalopathy, fibrosis and liver cancer [64]. Therefore, it's urgent to explore and develop effective hepatopro-tective candidates. Many works have indicated that dioscin has potent hepatopro- tective effects through regulating multiple targets and biological processes (Fig. 3).

3.1.1. Protective effects and mechanism of dioscin against acute liver damage

Some chemicals and drugs including CCl₄, lipopolysaccharide (LPS), ANIT, DMN, acetaminophen (APAP) and ethanol have been widely used to establish experimental models for estimating the activities of medicines against acute liver injury. Extensive researches have proved that dioscin can reduce serum levels of alanine aminotran- sferase (ALT), aspartate transaminase (AST), and improve acute liver injury.

Cytochrome P450 2E1 (CYP2E1) can cause ROS production to induce hepatic oxidative stress, inflammation and apoptosis, and finally lead to cell necrosis and tissue damage. Lu, B.N. et al., have found that dioscin shows significant protective effect against CCl₄-induced acute liver damage in mice, and the proteins including heat shock protein 5 (HSPA5), annexin A6 (ANXA6), isovaleryl-CoA dehydrogenase (IVD), ribosomal protein S6 (RPS6), cytoglobin (Cygb), and nucleoside diphosphate kinase A (NDPK-A) may be the drug targets based on proteomics assay [65]. Lu B.N. et al., have further found that dioscin can inhibit inflammation, apoptosis, necrosis and lipid peroxidation [66]. In addition, Zhao, X.M. et al., have investigated the effect of dioscin against APAP-induced liver injury, and found that dioscin can improve AST release, mitochondrial dysfunction and apoptosis in mice. Some differentially expressed proteins including sulfite oxidase (Suox), cytoskeletal 18 (Krt18), regucalcin (Rgn), PRDX1, malate dehydrogenase (MDH) and purine nucleoside phosphorylase (PNP) may be the drug targets of the compound against APAP-induced liver injury, which should be considered as biomarkers for investigating acute liver injury [67].

LPS can induce serious sepsis, activate immunological system and macrophages. Yao, H. et al., have showed that dioscin has potent action against LPS-induced liver injury through inhibiting toll-like receptor 4 (TLR4) inflammatory signaling pathway [68]. In addition, Yao, H.



Fig. 3. Dioscin possesses hepatoprotective effects through regulating multiple targets and biological processes including steatosis, inflammatory response, oxidative stress, apoptosis, TGF-B/Smads, autophagy and MAPKs pathways, α -SMA, α smooth muscle actin; ALT, alanine aminotransferase; AST, aspartate transaminase; Bax, Bcl2-associated X; Bcl-2, B-cell lymphoma-2; COL1 A1, collagen type I alpha 1; COL3A1, collagen, type III, alpha 1; HMGB-1, high mobility group box-1 protein; HO-1, heme oxygenase-1; IL, interleukin; IRF9, interferon regulatory factor 9; ITGA5, inhibiting integrin alpha-5; MAPKs, mitogen-activated protein kinases; MyD88, myeloid differentiation primary response 88; NAFLD, nonalcoholic fatty liver disease; Nrf2, nuclear factor (erythroid-derived 2)like 2; ROS, reactive oxygen species; Sirt1, sirtuin 1; SDC-4, syndecan-4; TGF-β1, transforming growth factor-β; TLR4, tolllike receptor 4; TNF-a, tumor necrosis factor-a; TRAF6, TNF receptor associated factor 6.

et al., have also proved that dioscin has potent effects against ANITinduced intrahepatic cholestasis through regulating apoptosis and ROS [69]. The study performed by Zhang, A.J. et al., have proved that dioscin can ameliorate cholestasis by restoring hepatic transporter expressions [70]. DMN is a potent hepatotoxin, mutagen and carcinogen, and thioacetamide (TAA) is another classic liver toxic chemical, which have been used to establish experimental models. The works of Zhang, W.X. et al., have indicated that dioscin exhibits protective effect against DMN-induced liver insult through inhibiting TLR4 inflammatory pathway and activating sirtuin 1 (Sirt1) antioxidant signaling pathway [71]. Zheng, L.L. et al., have investigated the protective effect of dioscin against TAA-induced acute liver injury, and dioscin can notably inhibit oxidative stress and inflammation in liver via adjusting farnesoid X receptor (FXR) signal pathway [72]. In addition, alcohol abuse can cause alcoholic liver diseases with the morphological features of steatosis, hepatitis, fibrosis and ultimately cirrhosis. In the paper of Xu, T.T. et al., the actions of dioscin against alcoholic liver disease have been investigated, and dioscin can notably improve liver steatosis, and decrease ALT and AST levels via ameliorating ethanol-induced oxidative stress, mitochondrial function, inflammatory cytokine production and apoptosis [73]. During I/R injury, activated Kupffer cells can produce massive ROS. Our study has also proved that the protective effects of dioscin against liver I/R injury may be through inhibiting inflammatory cytokines, oxidative stress, apoptosis and necrosis [74].

3.1.2. Protective effects and mechanism of dioscin against chronic liver damage

Overproduction of extracellular matrix (ECM) in liver can damage hepatic architecture and normal function, and finally cause fibrosis, cirrhosis and liver cancer. Molecular mechanistic researches have indicated that hepatic stellate cells (HSCs) activation is an important effector in the process of hepatic fibrosis [75]. Briefly, HSCs can be activated from quiescent cells to myofibroblast-like cells. Alcohol abuse, obesity, hepatitis, biliary disease, metabolic disorders, drug-induced chronic hepatic diseases, and continued exposure to chemicals or poisons can cause liver fibrosis [76,77].

Multiple studies have proved that dioscin can notably improve hepatic fibrosis, and inhibit cell viabilities of HSC-T6, LX-2 and primary rat HSCs. Dioscin can reduce HSCs activation and collagen accumulation based on the decreased levels of TGF-B1, α-SMA, collagen and vimentin, in vivo, the hepatic fibrosis models induced by CCl₄, alcoholic, TAA, bile duct ligation (BDL) and DMN have been used to test the actions of dioscin [78-82]. The results have confirmed that dioscin exerts anti-fibrotic activities with the improved body weights, serum AST, ALT and hydroxyproline levels via modulating multiple signaling pathways. TGF-B1 can phosphorylate downstream receptor-activated Smads signal [83]. Wnt/β-catenin pathway participates in HSCs activation [84]. Oxidative stress and inflammation also widely participate in HSCs activation and matrix degradation [85]. Among them, TLR4/ MyD88 signaling exhibits potent action against inflammatory response by decreasing NF-kB, interleukin (IL)-1, IL-6 and tumor necrosis factor- α (TNF- α) levels [86]. Nrf2/Sirt1 signaling can reduce p38 MAPK phosphorylation, and decrease the levels of collagen type I alpha 1 (COL1 A1), collagen type III alpha 1 (COL3A1), α-SMA and fibronectin through inhibiting ROS production [87]. Our previous studies have proved that the effects of dioscin against hepatic fibrosis may be via inhibiting TGF-\u00df1/Smads, Wnt/\u00bf-catenin and TLR4/MyD88 signal pathways and activating Nrf2/Sirt1 signal pathway [78-82].

As shown in Fig. 4A, two-dimensional differential in-gel electrophoresis technology (2D-DIGE) has been used to identify the differentially expressed proteins caused by dioscin in rats, and ten new biomarkers including protein disulfide isomerase A3 (PDIA3), seleniumbinding protein 1 (SBP1), glutamine synthetase (GSS), senescence marker protein 30 (RGN), hemopexin, keratin 8, keratin 18, vimentin,



Fig. 4. Dioscin exerts anti-liver fibrosis effects via affecting mutiple signaling pathways and biological processes. (A) Representative 2D-gel image marked with a total of 48 differentially expressed protein spots with red color for indicating the up-regulation of protein expressions and green color for showing the down-regulation of protein expressions. (B) The network of the differentially expressed proteins found by the iTRAQ method in HSC-T6 cells treated by dioscin. (C) Classification of the differentially expressed proteins found by SILAC proteomics in LX-2 cells treated by dioscin according to biological process. (D) The binding mode of dioscin and ITGA5 protein. These images are obtained from our previous results published in the References [81,87,88]. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article).

annexin A5 (ANXA5) and dermatopontin (DPT) have been found [81]. Syndecan-4 (SDC-4) plays a crucial role in effects of dioscin on HSC-T6 cell adhesion and migration [87], and the network of the differentially expressed proteins analyzed by the String 9.1 software is shown in Fig. 4B. In addition, as shown in Fig. 4C, Xu L.N. et al., have adopted the method of stable isotope labeling with amino acids in cell culture (SILAC) and found that dioscin can alleviate hepatic fibrosis via affecting multi-biological processes [88]. Moreover, dioscin can reduce collagen synthesis in LX-2 cells, and integrin alpha-5 (ITGA5) is a potent drug-target of dioscin based on molecular docking test (Fig. 4D).

3.1.3. Dioscin promotes liver regeneration

Currently, liver regeneration is a critical process after partial hepatectomy [89]. Many studies have suggested that Notch1 and Jagged1 are critical players in liver proliferation [90]. Jagged1 can activate Notch1 signal pathway, and cause the release of Notch1 intracellular domain (NICD1). Then, NICD1 translocates into nucleus and results in the transcription of proliferation-related genes. Dioscin can activate Notch1 /Jagged1 pathway to promote liver proliferation [91].

3.2. Protective effects and mechanism of dioscin on cardiovascular and cerebral vessels

Although arterial bypass and other surgical operations have been widely applied, myocardial I/R injury has become a primary factor in the treatment of coronary heart disease [92]. In the study of Qin, J. *et. al.*, the apoptosis of H9c2 cells induced by myocardial I/R can be prevented by dioscin via adjusting apoptotic pathway [93]. Doxorubicin is

an efficient chemotherapeutic drug which has been widely used in clinical treatment. However, the acute and chronic cardiotoxicities including arrhythmia, tachycardia, hypotension and refractory late-onset cardiomyopathy limit its clinical application. Zhao, L.S. et al., have found that dioscin can alleviate doxorubicin-induced cardiotoxicity via inhibiting oxidative stress through modulating miR-140-5p signaling pathway [94]. Cerebral I/R injury is one challenging clinical problem in the treatment of acute ischemic stroke. Some pathological mechanisms associated with cerebral I/R injury including oxidative stress, inflammation, and apoptosis have been implicated [95]. Dioscin shows protective effect against cerebral I/R injury in virto and in vivo [96,97] via inhibiting the expression level and nuclear translocation of high mobility group box-1 protein (HMGB-1). Moreover, dioscin can decrease transcriptional activities of NF-kB to suppress inflammatory response via inhibiting TLR4 signal. Also, small interfering RNA (siRNA) and over-expression experiments have confirmed that diosicn processes neuroprotective effect via regulating HMGB-1/TLR4 signal pathway. Atherosclerosis, a chronic disease with the accumulation of lipids and fibrous elements in large arteries, is the main cause of coronary artery disease. Monocyte-endothelial adhesion is an early step in the process of atherosclerosis. Dioscin can decrease monocyte adhesion via downregulating the expression levels of vascular cell adhesion molecule-1 (VCAM-1) and intercellular adhesion molecule 1 (ICAM-1), and inhibit endothelial lipase (EL) expression and NF-KB signal pathway [98].

3.3. Nephroprotective effects and mechanism of dioscin

Acute kidney injury associated with high morbidity and mortality is one common disease in clinical. Hyperuricemia, kidney transplantation and renal surgery-induced renal I/R injury, endotoxic (LPS) shock and chemotherapy are the common causes of kidney injury [99]. Renal I/R injury can cause the clinical syndrome of delayed graft function after transplantation. Dioscin exerts protective effects against renal I/R injury through inhibiting inflammatory response via down-regulating HSP70/TLR4 signaling pathway [100]. Dioscin can also decrease BUN and Cr levels, and ameliorate oxidative stress in vivo and in vitro models of LPS-induced inflammatory kidney injury via increasing miRNA let-7i level and thereby decreasing TLR4 inflammatory pathway [101]. Cisplatin, an inorganic platinum compound with broad-spectrum activities against tumours, can cause serious nephrotoxicities, which limit its clinical applications. The works of Zhang, Y.M, et al., have found that dioscin can attenuate cisplatin-induced renal injury in rats and mice via down- regulating miR-34a level, and then activacting Sirt1 signaling pathway [102]. Furthermore, Zhang, Y.M. et al., have also investigated the action of dioscin against doxorubicin-induced nephrotoxicity, and found that dioscin can markedly attenuate cell injury and nephrotoxicity in rats through inhibiting ROS level [103] via activating FXR siganling pathway. In addition, some food additives can also cause renal injury, and recent studies have shown excessive consumption of fructose can promote acute and chronic kidney injury. The works of Qiao, Y.J. et al., have indicated that dioscin can significantly reduce fructose-induced renal injury through decreasing the levels of Cr, BUN, and rehabilitating histopathological changes [104] via adjusting sirtuin 3 (Sirt3)-mediated signaling pathway.

3.4. Pulmonary protective effects and mechanism of dioscin

Airway mucus plays important roles against noxious chemicals, invading pathogenic microorganisms and particles. Abnormality in mucins always can cause severe pulmonary diseases. Lee, H.J. et al., have investigated the effect of dioscin on airway epithelial NCI-H292 cells, and found that dioscin can inhibit the gene expression and production of MUC5AC mucin induced by epidermal growth factor (EGF) or phorbol 12-myristate 13-acetate (PMA) [105]. Silicosis is caused by the inhalation of respirable crystalline silica dust, which can result in pulmonary fibrosis and even respiratory failure [106]. Li, C. et al., have found that dioscin can postpone crystalline silica-induced pulmonary fibrosis and exert protective actions in mice via inhibiting the secretion of pro-inflammation and pro-fibrotic cytokine [107]. In addition, dioscin can also inhibit signal-regulating kinase 1 (ASK-1)-p38/JNK pathway and Smad3 phosphorylation in fibroblast. Furthermore, Zeng, H.Q. et al., have investigated the protective effect and underlying mechanism of dioscin against acute lung injury induced by LPS, and found that dioscin can significantly decrease total number of alveolar macrophages and water content of lung in LPS-treated mice through HSP70/TLR4 inflammatory signaling pathway [108].

3.5. Protective effects and mechanism of dioscin against gastric I/R injury

Many clinical conditions including ischemia gastrointestinal disease, vascular rupture or surgery and hemorrhagic shock can cause gastric I/R injury. Dioscin can attenuate H/R insult in GES-1 cells and gastric I/R injury in rats through inhibiting NF- κ B and AP-1 transcriptional activities, and pro-inflammatory cytokine responses via inhibiting PKC/ERK1/2 signal pathway [109].

4. Anti-metabolic diseases effects and mechanism of dioscin

Metabolic diseases are caused by a group of risk factors including osteoporosis, diabetes, obesity and insulin resistance [110,111]. Nonalcoholic fatty liver disease (NAFLD) is a clinicopathologic spectrum of liver pathologies with excessive accumulation of fat in liver [112]. The activity of dioscin against NAFLD has been investigated by Xu, L.N. et al., and dioscin exerts obvious protective effects against NAFLD through affecting multiple drug targets and signal pathways [113]. Using iTRAQ labeling coupled with nano-LC-TOFMS/MS analysis, 22 proteins including enoyl-CoA delta isomerase 1 (Eci1), acyl-CoA dehydrogenase short chain (ACADS), acyl-CoA synthetases long-chain 5 (ACLS5), aldehyde dehydrogenase 7 family member A1 (ALDH7A1), aldehyde dehydrogenase 2 (ALDH2) and cytochrome P450 4 A2 (CYP4 A2) maybe the drug tragets of dioscin against NAFLD. Furthermore, obesity is featured by an excessive storage of body fat associated with a great number of metabolic complications. Liu, M. et al., have proved that dioscin can alleviate liver lipid accumulation symptoms, increase oxygen consumption and energy expenditure in high-fat dietinduced C57BL/6J mice and ob/ob mice via inhibiting oxidative damage, inflammation and MAPK phosphorylation levels [114]. Poudel, B. et al., have also showed that dioscin can inhibit adipocyte differentiation in 3T3-L1 cells and decrease weight gain in mice against highfat diet-induced obesity [115].

Osteoporosis can be caused by excessive bone resorption, and correcting the imbalance between bone resorption and formation is effective method for treating osteoporosis [116]. Dioscin can inhibit the receptor activator of NF-KB ligand (RANKL) -mediated osteoclast differentiation and bone resorption in vitro. Furthermore, in vivo study has verified the protective activity of dioscin in LPS-induced osteolytic mice modle by inhibiting NF-κB signaling pathway and nuclear factor of activated T-cells 1 (NFATc1) transcriptional activity [117]. Our previous paper has also found that dioscin can notably improve the biochemical indexes and microarchitecture in ovariectomy (OVX)-induced animal models through promoting osteoblastogenesis and inhibiting osteoclastogenesis [118]. Osteoblasts are also regulated by hormonal and local factors including Wnt/Lrp5/β-catenin and estrogen receptor (ER) signaling pathways. Zhang, C.F. et al., have found that dioscin can increase the proliferation and differentiation of osteoblasts by up-regulating ER and Wnt/Lrp5/ β-catenin signaling pathways [119]. In addition, Zhun, C. et al., have identified that dioscin can promote MC3T3-E1 cell proliferation and differentiation by regulating autophagy [120]. Dioscin maybe a novel and potent candidate for the treatment of osteoporosis.

Hyperuricemia with high level of blood uric acid has been recognized as a risk factor of hypertension, gout, cardiovascular disease,



Fig. 5. Pharmacological activities of dioscin reported in recent years. In recent years, dioscin has reached more and more attention with its potent effects to treat multiple organ injuries and metabolic diseases as well as its anti-cancer activities through adjusting multiple targets and multiple signals. TLR4, toll-like receptor 4; MyD88, myeloid differentiation primary response 88; TRAF6, TNF receptor associated factor 6; IKKs, IkB kinases; JNK, c-Jun N-terminal kinase; AP-1, activating protein-1; NF-κB, nuclear factor κB; STAT3, signal transducer and activator of transcription 3; FXR, farnesoid X receptor; Nrf2, nuclear factor (erythroid-derived 2)-like 2; Sirt1, sirtuin 1; TGF-β, transforming growth factor-β.

diabetes and chronic kidney disease [121,122]. Dioscin can significantly decrease serum uric acid and Cr levels, and reduce renal pathological lesions caused by hyperuricemia in mice [123]. Urate transporter 1 (URAT1) has been identified to be involved in renal transportion of uric acid. Moreover, ATP-binding cassette subfamily G member 2 (ABCG2) is an efficient urate exporter to mediate renal and intestinal urate excretion, and controls serum uric acid levels. Dioscin can decrease serum level of uric acid through inhibiting URAT1 and promoting ABCG2.

Furthermore, type 2 diabetes is one common metabolic disorder. However, current therapies have some undesirable side effects including weight gain, dropsy, drug resistance and high rates of econdary failure [124]. Hence, it is necessary to disover more effective antidiabetic drugs with less side effects. Yu, H. et al., have evaluated the efficacy of the total saponins from *Dioscorea nipponica* Makino (TSDN) against T2DM [125]. The results showed that TSDN can significantly decrease the fasting blood glucose, ameliorate the levels of oral glucose and insulin tolerance test, and markedly increase body weight and serum insulin. Via adjusting MAPKs, NF- κ B, peroxisome proliferatoractivated receptor γ (PPAR γ) and glycogen synthase kinase-3beta (GSK-3 β) pathways.

5. Other pharmacological activities of dioscin

Adenovirus, a common cause of diarrhea, accounts for about 10% of acute respiratory infections in young children [126]. Vesicular stomatitis virus (VSV), a member of Rhabdoviridae family, can lead to oral diseases including mucosal vesicles and ulcers in mouth [127]. Hepatitis B virus (HBV) belongs to Hepadnaviridae family, and HBV infection can cause liver disease and hepatocellular carcinoma [128]. Up to now, there have no efficient antiviral drugs to treat adenoviral infections, VSV and HBV. According to the research of Liu, C.H. et al., antiviral effects of dioscin have been tested and dioscin can block the initial stage of adenovirus infection in 293 cells and affect the host cell's response for viral infection [129]. Moreover, dioscin exerts anti- VSV infection action prior to infection. In addition, dioscin has potent inhibitory effect on the secretion of Hepatitis B e-antigen (HBeAg) and Hepatitis B surface antigen (HBsAg) in HepG2 2.2.15 cells. Candida albicans can lead to candidiasis, which is the primary cause of notable morbidity and mortality in immunocom- promised patients [130]. In the study of Cho, J.Y. et al, dioscin exerts the effect of membrane disruption via inducing morphological change and rhodamine leakage of the giant unilamellar vesicles (GUVs) [131]. Rheumatoid arthritis is a chronic autoimmune disease associated with serious cardiovascular complications. Guo, Y.C. et al., have detected the therapeutic effect of dioscin on collagen-induced arthritis (CIA), and found that dioscin can regulate the proportion of Th1/Th2 cells [132]. In addition, dioscin can also balance the Th17 and Treg cell specific transcription factor retinoic acid receptorrelated orphan receptor gamma (RORy) and forkhead box P3 (Foxp3), and inhibit inflammatory reaction in CIA [133-135]. Growth hormone is a peptide hormone consisting 191 amino acids, which has been used to treat dwarfism and some diseases including obesity, osteoporosis and aging; and Lee, H.Y. et al., have also found that dioscin can induce the release of growth hormone in rats [136].

6. Clinical application

Currently, there have no clinical studies about dioscin because it is only one lead compound not a drug that can not be used on humans. However, some traditional Chinese medicines (TCMs) containing dioscin as the major active compound have been widely used in clinical. Liuwei Dihuang decoction (LW) is one of the most famous TCM formulae, which has been used to treat some kinds of diseases including cancer, neurosis, neurasthenia, dementia, Parkinson's disease, osteoporosis, diabetics, hypertension, nephritis, and thrombocytopenic purpura with the characteristic features of kidney Yin deficiency since Song Dynasty probably 900 years ago [137,138]. In addition, another popular formula-Di'ao Xinxue kang (Di'ao XXK) prepared from Dioscorea nipponica Makino has anti-coronary disease, anti- angina pectoris, and anti-myocardial ischemia effects [139], which has been used for many years in China to treat coronary heart disease [140]. Furthermore, Dioscornin Tablet contains some kinds of saponins that are extracted from Dioscorea nipponica, which has been used to treat Ca²⁺ overload in cardiomyocytes of coronary heart disease in clinical [141]. Therefore, dioscin maybe a potential drug candidate for the further clinical trials.

7. Conclusions

In conclusion, dioscin has attracted a great attention worldwide, and hundreds of papers have been reported to describe the pharmacological activities and mechanisms of this natural product (Fig. 5). Most of these researchers have described the mechanistic effects of dioscin on oxidative stress, inflammation, apoptosis, autophagy, lipidlowering and immunization on multiple body systems. This review paper addressed the effects of dioscin on multiple targets and signal pathways in a variety of diseases. Dioscin exerts the potent anti-cancer activities through regulating DNA damage, cell cycle arrest, apoptosis, autophagy, ROS level and Ca²⁺ release, and protects multiple organs injuries via the inhibiton of inflammation, apoptosis, necrosis, lipid peroxidation and mitochondrial dysfunction. Moreover, dioscin has the abilities against metabolic diseases through affecting lipid accumulation, oxidative damage, autophagy and glucose metabolism. Meanwhile, dioscin can also inhibit viruses and bacteria, regulate immunization and induce growth hormone. These years, we have done many works on dioscin, and found that these diverse therapeutic actions may involve in the TLR4/MyD88, Sirt1/Nrf2, FXR and TGF- β /Smads signal pathways. However, the currently available pharmacodynamic and mechanistic data are derived from in vitro cell experiments and in vivo rodent models, which require additional translator work before the human-specific clinical conclusions to be drawn.

Competing financial interests

The authors declare no competing financial interests.

Acknowledgments

This work was financially supported byNational Natural Science Foundation of China (No. 81872921 and No. 81603178), the Key Research and Development Project of Liaoning Province (No. 2017225090), the Science and Technology Innovation Project of Dalian, China (No. 2018J128N083), the Project of Liaoning BaiQianWan Talents Program (No. 2015-65), the Special Grant for Translational Medicine, Dalian Medical University (No. 2015004) and the Basic Scientific Research Projects of Liaoning University (No. LF2017010).

References

- Institute for Health Metrics and Evaluation. GBD 2015 Protocol: global burden of diseases, injuries, and risk factors. 2015; Available at: http://www.healthdata. org/gbd/about/protocol.
- [2] J.J. Lu, W. Pan, Y.J. Hu, Y.T. Wang, Multi-target drugs: the trend of drug research and development, PloS One 2012 (7) (2012) e40262.
- [3] N.W. Morrell, S.L. Archer, A. Defelice, S. Evans, M. Fiszman, T. Martin, et al., Anticipated classes of new medications and molecular targets for pulmonary arterial hypertension, Pulm. Circ. (3) (2013) 226–244.
- [4] R. Yuan, Y. Lin, Traditional Chinese medicine: an approach to scientific proof and clinical validation, Pharmacol. Ther. (86) (2000) 191–198.
- [5] N. Zhao, J. Li, L. Li, X.Y. Niu, M. Jiang, X.J. He, et al., Molecular network-based analysis of guizhi-shaoyao-zhimu decoction, a TCM herbal formula, for treatment of diabetic peripheral neuropathy, Acta Pharmacol. Sin. (36) (2015) 716–723.
- [6] M. Dashtdar, M.R. Dashtdar, B. Dashtdar, K. Kardi, M.K. Shirazi, The concept of wind in traditional Chinese medicine, J. Pharmacopuncture (19) (2016) 293–302.
- [7] X. Cheng, X. Su, X. Chen, H. Zhao, C. Bo, J. Xu, et al., Biological ingredient analysis of traditional Chinese medicine preparation based on high-throughput sequencing: the story for Liuwei Dihuang Wan, Sci. Rep. (4) (2014) 5147.
- [8] W. Zhou, X. Cheng, Y. Zhang, Effect of Liuwei Dihuang decoction, a traditional Chinese medicinal prescription, on the neuroendocrine immunomodulation network, Pharmacol. Ther. 162 (2016) 170–178.
- [9] Y. Jia, C. Chen, C.S. Ng, S.W. Leung, Meta-analysis of randomized controlled trials on the efficacy of Di'ao Xinxuekang capsule and isosorbide dinitrate in treating angina pectoris, Evid. Based Complement. Alternat. Med. (2012) (2012) 904147.
- [10] Y. Yu, S. Hu, G. Li, J. Xue, Z. Li, X. Liu, et al., Comparative effectiveness of Di'ao Xin Xue Kang capsule and compound Danshen tablet in patients with symptomatic chronic stable angina, Sci. Rep. (4) (2014) 7058.
- [11] Y.N. Tang, Y.X. Pang, X.C. He, Y.Z. Zhang, J.Y. Zhang, Z.Z. Zhao, et al., UPLC-QTOF- MS identification of metabolites in rat biosamples after oral administration of *Dioscorea saponins*: a comparative study, J. Ethnopharmacol. 165 (2015) 127–140.
- [12] L. Xu, Y. Wei, J.Y. Peng, Advances in study of dioscin–a natural product, Zhongguo Zhong Yao Za Zhi 40 (2015) 36–41.
- [13] Y. Zhou, Y. Su, Z. Zeng, X. Huang, P. Zhou, Exploration on compatibility between Shanyao and top grade medicinals in the Shen Nong Ben Cao Jing, World Sci. Technol. 15 (2013) 2060–2062.
- [14] R. Siegel, J. Ma, Z. Zou, A. Jemal, Cancer statistics, 2014, CA. Cancer J. Clin. 64 (2015) 9–29.
- [15] N.M. Dimitriou, G. Tsekenis, E.C. Balanikas, A. Pavlopoulou, M. Mitsiogianni, T. Mantso, et al., Gold nanoparticles, radiations and the immune system: current insights into the physical mechanisms and the biological interactions of this new alliance towards cancer therapy, Pharmacol. Ther. 178 (2017) 1–17.
- [16] F. Cardoso, N. Harbeck, L. Fallowfield, S. Kyriakides, E. Senkus, Locally recurrent or metastatic breast cancer: ESMO clinical practice guidelines for diagnosis, treatment and follow-up, Ann. Oncol. 23 (Suppl. 7) (2012) vii11–19.

- [17] P. Aumsuwan, S.I. Khan, I.A. Khan, Z. Ali, B. Avula, L.A. Walker, et al., The anticancer potential of steroidal saponin, dioscin, isolated from wild yam (*Dioscorea villosa*) root extract in invasive human breast cancer cell line MDA-MB-231 in vitro, Arch. Biochem. Biophys. 591 (2016) 98–110.
- [18] P. Aumsuwan, S.I. Khan, I.A. Khan, L.A. Walker, A.K. Dasmahapatra, Gene expression profiling and pathway analysis data in MCF-7 and MDA-MB-231 human breast cancer cell lines treated with dioscin, Data Brief 8 (2016) 272–279.
- [19] E.A. Kim, J.H. Jang, Y.H. Lee, E.G. Sung, I.H. Song, J.Y. Kim, et al., Dioscin induces caspase-independent apoptosis through activation of apoptosis-inducing factor in breast cancer cells, Apoptosis 19 (2014) 1165–1175.
- [20] H. Glavinas, P. Krajcsi, J. Cserepes, B. Sarkadi, The role of ABC transporters in drug resistance, metabolism and toxicity, Curr. Drug Deliv. 1 (2014) 27–42.
- [21] M.G. Garcia, L.D. Alaniz, R.I. Cordo Russo, E. Alvarez, S.E. Hajos, PI3K/Akt inhibition modulates multidrug resistance and activates NF-kappaB in murine lymphoma cell lines, Leuk. Res. 33 (2009) 288–296.
- [22] L. Wang, C. Wang, Y. Jia, Z. Liu, X. Shu, K. Liu, Resveratrol increases anti- proliferative activity of bestatin through downregulating P-glycoprotein expression via inhibiting PI3K/Akt/mTOR pathway in K562/ADR cells, J. Cell Biochem. 117 (2016) 1233–1239.
- [23] C. Wang, X. Huo, L. Wang, Q. Meng, Z. Liu, Q. Liu, et al., Dioscin strengthens the efficiency of adriamycin in MCF-7 and MCF-7/ADR cells through autophagy induction: more than just down-regulation of MDR1, Sci. Rep. 6 (2016) 28403.
- [24] H.J. Lee, I.C. Song, H.J. Yun, D.Y. Jo, S. Kim, CXC chemokines and chemokine receptors in gastric cancer: from basic findings towards therapeutic targeting, World J. Gastroenterol. 20 (2014) 1681–1693.
- [25] M. Hu, L. Xu, L. Yin, Y. Qi, H. Li, Y. Xu, et al., Cytotoxicity of dioscin in human gastric carcinoma cells through death receptor and mitochondrial pathways, J. Appl. Toxicol. 33 (2013) 712–722.
- [26] D.O. Morgan, Principles of CDK regulation, Nature 374 (1995) 131-134.
- [27] L.L. Gao, F.R. Li, P. Jiao, S.T. Yao, H. Sang, Y.H. Si, Apoptosis of human ovarian cancer cells induced by *Paris chinensis* dioscin via a Ca²⁺-mediated mitochondrion pathway, Asian Pac. J. Cancer Prev. 12 (2011) 1361–1366.
- [28] M. Liu, L. Xu, L. Yin, Y. Qi, Y. Xu, X. Han, et al., Potent effects of dioscin against gastric cancer *in vitro* and in vivo, Phytomedicine 23 (2016) 274–282.
- [29] J.M. Silva, D.S. Perez, J.R. Pritchett, M.L. Halling, H. Tang, D.I. Smith, Identification of long stress-induced non-coding transcripts that have altered expression in cancer, Genomics 95 (2010) 355–362.
- [30] T. Ma, R.P. Wang, X. Zou, Dioscin inhibits gastric tumor growth through regulating the expression level of lncRNA HOTAIR, BMC Complement. Altern. Med. 16 (2016) 383.
- [31] C. Fitzmaurice, D. Dicker, A. Pain, H. Hamavid, M. Moradi-Lakeh, M.F. MacIntyre, et al., The global burden of cancer 2013, JAMA Oncol. 1 (2015) 505–527.
- [32] K. Mauer, R. O'Kelley, N. Podda, S. Flanagan, S. Gadani, New treatment modalities for hepatocellular cancer, Curr. Gastroenterol. Rep. 17 (2015).
- [33] M.J. Hsieh, S.F. Yang, Y.S. Hsieh, T.Y. Chen, H.L. Chiou, Autophagy inhibition enhances apoptosis induced by dioscin in huh7 cells, Evid. Based Complement. Alternat. Med. 2012 (2012) 134512.
- [34] G. Zhang, X. Zeng, R. Zhang, J. Liu, W. Zhang, Y. Zhao, et al., Dioscin suppresses hepatocellular carcinoma tumor growth by inducing apoptosis and regulation of TP53, BAX, BCL2 and cleaved CASP3, Phytomedicine 23 (2016) 1329–1336.
- [35] B.T. Sun, L.H. Zheng, Y.L. Bao, C.L. Yu, Y. Wu, X.Y. Meng, et al., Reversal effect of dioscin on multidrug resistance in human hepatoma HepG2/adriamycin cells, Eur. J. Pharmacol. 654 (2011) 129–134.
- [36] P.H. Alizai, L. Bertram, A. Fragoulis, C.J. Wruck, D.C. Kroy, U. Klinge, et al., *In vivo* imaging of antioxidant response element activity during liver regeneration after partial hepatectomy, J. Surg. Res. 206 (2016) 525–535.
- [37] Y. Wang, Q.Y. He, J.F. Chiu, Dioscin induced activation of p38 MAPK and JNK via mitochondrial pathway in HL-60 cell line, Eur. J. Pharmacol. 735 (2014) 52–58.
- [38] L. Wang, Q. Meng, C. Wang, Q. Liu, J. Peng, X. Huo, et al., Dioscin restores the activity of the anticancer agent adriamycin in multidrug-resistant human leukemia K562/adriamycin cells by down-regulating MDR1 via a mechanism involving NFkappaB signaling inhibition, J. Nat. Prod. 76 (2013) 909–914.
- [39] Y. Wei, Y. Xu, X. Han, Y. Qi, L. Xu, Y. Xu, et al., Anti-cancer effects of dioscin on three kinds of human lung cancer cell lines through inducing DNA damage and activating mitochondrial signal pathway, Food Chem. Toxicol. 59 (2013) 118–128.
- [40] M.J. Hsieh, T.L. Tsai, Y.S. Hsieh, C.J. Wang, H.L. Chiou, Dioscin-induced autophagy mitigates cell apoptosis through modulation of PI3K/Akt and ERK and JNK signaling pathways in human lung cancer cell lines, Arch. Toxicol. 87 (2013) 1927–1937.
- [41] W.C. Lim, H. Kim, Y.J. Kim, K.C. Choi, I.H. Lee, K.H. Lee, et al., Dioscin suppresses TGF-beta1-induced epithelial- mesenchymal transition and suppresses A549 lung cancer migration and invasion, Bioorg. Med. Chem. Lett. 27 (2017) 3342–3348.
- [42] Y.C. Wang, D.W. Wu, T.C. Wu, L. Wang, C.Y. Chen, H. Lee, et al., Dioscin overcome TKI resistance in EGFR-mutated lung adenocarcinoma cells via down- regulation of tyrosine phosphatase SHP2 expression, Int. J. Biol. Sci. 14 (2018) 47–56.
- [43] Y.S. Kim, E.A. Kim, K.G. Park, S.J. Lee, M.S. Kim, H.Y. Sohn, T.J. Lee, Dioscin sensitizes cells to TRAIL-induced apoptosis through downregulation of c-FLIP and bcl-2, Oncol. Rep. 28 (2012) 1910–1916.
- [44] G. Zhang, J. Xiao, H. Shao, X. Lai, P. Qiu, Y. Wu, et al., Effects of dioscin on the gap junction function in 786-0 cell of human renal carcinoma, Chin. Pharmacolo-gical Bull. 28 (2012) 778–782.
- [45] Y.J. Jeong, J.Y. Lee, J. Park, S.N. Park, An inhibitory mechanism of action of a novel syringic-acid derivative on alpha-melanocyte-stimulating hormone (alpha-MSH)-induced melanogenesis, Life Sci. 191 (2017) 52–58.
- [46] J. Zhou, T. Ren, Y. Li, A. Cheng, W. Xie, L. Xu, et al., Oleoylethanolamide inhibits

Pharmacological Research 137 (2018) 259-269

alpha-melanocyte stimulating hormone-stimulated melanogenesis via ERK, akt and CREB signaling pathways in B16 melanoma cells, Oncotarget 8 (2017) 56868–56879.

- [47] A. Nishina, K. Ebina, M. Ukiya, M. Fukatsu, M. Koketsu, M. Ninomiya, et al., Dioscin derived from *Solanum melongena* L. "Usukawamarunasu" Attenuates alpha-MSH-induced melanogenesis in B16 murine melanoma cells via downregulation of phospho-CREB and MITF, J. Food Sci. 80 (2015) H2354–2359.
- [48] Y. Kou, L. Ji, H. Wang, W. Wang, H. Zheng, J. Zou, et al., Connexin 43 upregulation by dioscin inhibits melanoma progression via suppressing malignancy and inducing M1 polarization, Int. J. Cancer 141 (2017) 1690–1703.
- [49] A.A. Shafi, A.E. Yen, N.L. Weigel, Androgen receptors in hormone-dependent and castration- resistant prostate cancer, Pharmacol. Ther. 140 (2013) 223–238.
- [50] X. Tao, L. Xu, L. Yin, X. Han, Y. Qi, Y. Xu, et al., Dioscin induces prostate cancer cell apoptosis through activation of estrogen receptor-beta, Cell Death Dis. 8 (2017) e2989.
- [51] L.L. Gao, F.R. Li, P. Jiao, M.F. Yang, X.J. Zhou, Y.H. Si, et al., Paris chinensis dioscin induces G2/M cell cycle arrest and apoptosis in human gastric cancer SGC-7901 cells, World J. Gastroenterol. 17 (2011) 4389–4395.
- [52] Z. Wang, Y. Cheng, N. Wang, D.M. Wang, Y.W. Li, F. Han, et al., Dioscin induces cancer cell apoptosis through elevated oxidative stress mediated by down- regulation of peroxiredoxins, Cancer Biol. Ther. 13 (2012) 138–147.
- [53] L. Si, L. Zheng, L. Xu, L. Yin, X. Han, Y. Qi, et al., Dioscin suppresses human laryngeal cancer cells growth via induction of cell-cycle arrest and MAPK- mediated mitochondrial-derived apoptosis and inhibition of tumor invasion, Eur. J. Pharmacol. 774 (2016) 105–117.
- [54] X. Zhao, X. Tao, L. Xu, L. Yin, Y. Qi, Y. Xu, et al., Dioscin induces apoptosis in human cervical carcinoma HeLa and SiHa cells through ROS-mediated DNA damage and the mitochondrial signaling pathway, Molecules 21 (2016).
- [55] L. Lv, L. Zheng, D. Dong, L. Xu, L. Yin, Y. Xu, et al., Dioscin, a natural steroid saponin, induces apoptosis and DNA damage through reactive oxygen species: a potential new drug for treatment of glioblastoma multiforme, Food Chem. Toxicol. 59 (2013) 657–669.
- [56] L. Si, L. Xu, L. Yin, Y. Qi, X. Han, Y. Xu, et al., Potent effects of dioscin against pancreatic cancer via miR-149-3P-mediated inhibition of the Akt1 signalling pathway, Br. J. Pharmacol. 174 (2017) 553–568.
- [57] Q. Zhou, W. Song, W. Xiao, Dioscin induces demethylation of DAPK-1 and RASSF-1alpha genes via the antioxidant capacity, resulting in apoptosis of bladder cancer T24 cells, Excli. J. 16 (2017) 101–112.
- [58] X. Song, Z. Wang, H. Liang, W. Zhang, Y. Ye, H. Li, et al., Dioscin induces gallbladder cancer apoptosis by inhibiting ROS-mediated PI3K/AKT signalling, Int. J. Biol. Sci. 13 (2017) 782–793.
- [59] S. Li, B. Cheng, L. Hou, L. Huang, Y. Cui, D. Xu, et al., Dioscin inhibits colon cancer cells' growth by reactive oxygen species-mediated mitochondrial dysfunction and p38 and JNK pathways, Anticancer Drugs 29 (2018) 234–242.
- [60] W. Liu, Z. Zhao, Y. Wang, W. Li, Q. Su, Q. Jia, et al., Dioscin inhibits stem-cell-like properties and tumor growth of osteosarcoma through Akt/GSK3/β-catenin signaling pathway, Cell Death Dis. 9 (2018) 343.
- [61] J. Fernandez, F. Bert, M.H. Nicolas-Chanoine, The challenges of multi-drug- resistance in hepatology, J. Hepatol. 65 (2016) 1043–1054.
- [62] P. Sarges, J.M. Steinberg, J.H. Lewis, Drug-induced liver injury: highlights from a review of the 2015 literature, Drug Saf. 39 (2016) 801–821.
- [63] M. Blachier, H. Leleu, M. Peck-Radosavljevic, D.C. Valla, F. Roudot-Thoraval, The burden of liver disease in Europe: a review of available epidemiological data, J. Hepatol. 58 (2013) 593–608.
- [64] D. Dong, S. Zhang, L. Yin, X. Tang, Y. Xu, X. Han, et al., Protective effects of the total saponins from *Rosa laevigata* Michx fruit against carbon tetrachloride- induced acute liver injury in mice, Food Chem. Toxicol. 62 (2013) 120–130.
- [65] B. Lu, L. Yin, L. Xu, J. Peng, Application of proteomic and bioinformatic techniques for studying the hepatoprotective effect of dioscin against CCl₄- induced liver damage in mice, Planta. Med. 77 (2011) 407–415.
- [66] B. Lu, Y. Xu, L. Xu, X. Cong, L. Yin, H. Li, et al., Mechanism investigation of dioscin against CCl₄-induced acute liver damage in mice, Environ. Toxicol. Pharmacol. 34 (2012) 127–135.
- [67] X. Zhao, X. Cong, L. Zheng, L. Xu, L. Yin, J. Peng, J. Dioscin, A natural steroid saponin, shows remarkable protective effect against acetaminophen-induced liver damage *in vitro* and *in vivo*, Toxicol. Lett. 214 (2012) 69–80.
- [68] H. Yao, C. Hu, L. Yin, X. Tao, L. Xu, Y. Qi, et al., Dioscin reduces lipopolysaccharide- induced inflammatory liver injury via regulating TLR4/MyD88 signal pathway, Int. Immunopharmacol. 36 (2016) 132–141.
- [69] H. Yao, Y. Xu, L. Yin, X. Tao, L. Xu, Y. Qi, et al., Dioscin protects ANIT-induced intrahepatic cholestasis through regulating transporters, apoptosis and oxidative stress, Front. Pharmacol. 8 (2017) 116.
- [70] A. Zhang, Y. Jia, Q. Xu, C. Wang, Q. Liu, Q. Meng, et al., Dioscin protects against ANIT-induced cholestasis via regulating oatps, Mrp2 and bsep expression in rats, Toxicol. Appl. Pharmacol. 305 (2016) 127–135.
- [71] W. Zhang, L. Yin, X. Tao, L. Xu, L. Zheng, X. Han, et al., Dioscin alleviates dimethylnitrosamine- induced acute liver injury through regulating apoptosis, oxidative stress and inflammation, Environ. Toxicol. Pharmacol. 45 (2016) 193–201.
- [72] L. Zheng, L. Yin, L. Xu, Y. Qi, H. Li, Y. Xu, et al., Protective effect of dioscin against thioacetamide- induced acute liver injury via FXR/AMPK signaling pathway in vivo, Biomed. Pharmacother. 97 (2017) 481–488.
- [73] T. Xu, L. Zheng, L. Xu, L. Yin, Y. Qi, Y. Xu, et al., Protective effects of dioscin against alcohol-induced liver injury, Arch. Toxicol. 88 (2014) 739–753.
- [74] X. Tao, X. Wan, Y. Xu, L. Xu, Y. Qi, L. Yin, et al., Dioscin attenuates hepatic ischemia-reperfusion injury in rats through inhibition of oxidative-nitrative stress, inflammation and apoptosis, Transplantation 98 (2014) 604–611.

- [75] S.L. Friedman, Mechanisms of hepatic fibrogenesis, Gastroenterology 134 (2008) 1655–1669.
- [76] W.Z. Mehal, J. Iredale, S.L. Friedman, Scraping fibrosis: expressway to the core of fibrosis, Nat. Med. 17 (2011) 552–553.
- [77] E. Mormone, J. George, N. Nieto, Molecular pathogenesis of hepatic fibrosis and current therapeutic approaches, Chem. Biol. Interact. 193 (2011) 225–231.
- [78] L. Gu, X. Tao, Y. Xu, X. Han, Y. Qi, L. Xu, et al., Dioscin alleviates BDL- and DMNinduced hepatic fibrosis via Sirt1/Nrf2-mediated inhibition of p38 MAPK pathway, Toxicol. Appl. Pharmacol. 292 (2016) 19–29.
- [79] M. Liu, Y. Xu, X. Han, L. Yin, L. Xu, Y. Qi, et al., Dioscin alleviates alcoholic liver fibrosis by attenuating hepatic stellate cell activation via the TLR4/MyD88/ NFkappaB signaling pathway, Sci. Rep. 5 (2015) 18038.
- [80] X. Zhang, X. Han, L. Yin, L. Xu, Y. Qi, Y. Xu, et al., Potent effects of dioscin against liver fibrosis, Sci. Rep. 5 (2015) 9713.
- [81] X. Zhang, L. Xu, L. Yin, Y. Qi, Y. Xu, X. Han, et al., Quantitative chemical proteomics for investigating the biomarkers of dioscin against liver fibrosis caused by CCl₄ in rats, Chem. Commun. (Camb.) 51 (2015) 11064–11067.
- [82] X. Zhang, Y. Xu, Y. Qi, X. Han, L. Yin, L. Xu, et al., Potent effects of dioscin against thioacetamide-induced liver fibrosis through attenuating oxidative stress in turn inhibiting inflammation, TGF-β/Smad and MAPK signaling pathways, J. Funct. Foods 16 (2015) 436–447.
- [83] Y. Inagaki, I. Okazaki, Emerging insights into transforming growth factor beta Smad signal in hepatic fibrogenesis, Gut 56 (2007) 284–292.
- [84] J.H. Cheng, H. She, Y.P. Han, J. Wang, S. Xiong, K. Asahina, et al., Wnt antagonism inhibits hepatic stellate cell activation and liver fibrosis, Am. J. Physiol. Gastrointest. Liver Physiol. 294 (2008) G39–49.
- [85] Y. Le, L. Chen, Y. Zhang, P. Bu, G. Dai, X. Cheng, Epalrestat stimulated oxidative stress, inflammation, and fibrogenesis in mouse liver, Toxicol. Sci. 163 (2017) 397–408.
- [86] T. Kawai, S. Akira, The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors, Nat. Immunol. 11 (2010) 373–384.
- [87] L. Yin, Y. Qi, Y. Xu, L. Xu, X. Han, X. Tao, et al., Dioscin inhibits HSC-T6 cell migration via adjusting SDC-4 expression: insights from iTRAQ-based quantitative proteomics, Front Pharmacol. 8 (2017) 665.
- [88] L. Xu, L. Yin, X. Tao, Y. Qi, X. Han, Y. Xu, et al., Dioscin, a potent ITGA5 inhibitor, reduces the synthesis of collagen against liver fibrosis: insights from SILAC- based proteomics analysis, Food Chem. Toxicol. 107 (2017) 318–328.
- [89] K. Azijli, B. Weyhenmeyer, G.J. Peters, S. de Jong, F.A. Kruyt, Non-canonical kinase signaling by the death ligand TRAIL in cancer cells: discord in the death receptor family, Cell Death Differ. 20 (2013) 858–868.
- [90] C. Köhler, A.W. Bell, W.C. Bowen, S.P. Monga, W. Fleig, G.K. Michalopoulos, Expression of notch-1 and its ligand jagged-1 in rat liver during liver regeneration, Hepatology 39 (2004) 1056–1065.
- [91] L. Xu, L. Gu, X. Tao, Y. Xu, Y. Qi, L. Yin, et al., Effect of dioscin on promoting liver regeneration via activating Notch1/Jagged1 signal pathway, Phytomedicine 38 (2018) 107–117.
- [92] Y. Wang, L. Jin, Y. Song, M. Zhang, D. Shan, Y. Liu, et al., Beta-arrestin 2 mediates cardiac ischemia-reperfusion injury via inhibiting GPCR-independent cell survival signalling, Cardiovasc. Res. 113 (2017) 1615–1626.
- [93] J. Qin, Y. Kang, Z. Xu, C. Zang, B. Fang, X. Liu, Dioscin prevents the mitochondrial apoptosis and attenuates oxidative stress in cardiac H9c2 cells, Drug Res. (Stuttg) 64 (2014) 47–52.
- [94] L. Zhao, X. Tao, Y. Qi, L. Xu, L. Yin, J. Peng, Protective effect of dioscin against doxorubicin-induced cardiotoxicity via adjusting microRNA-140-5p-mediated myocardial oxidative stress, Redox. Biol. 16 (2018) 189–198.
- [95] C.A. Molina, J. Alvarez-Sabin, Recanalization and reperfusion therapies for acute ischemic stroke, Cerebrovasc. Dis. 27 (Suppl. 1) (2009) 162–167.
- [96] X. Tao, X. Sun, L. Yin, X. Han, L. Xu, Y. Qi, et al., Dioscin ameliorates cerebral ischemia/reperfusion injury through the downregulation of TLR4 signaling via HMGB-1 inhibition, Free Radic. Biol. Med. 84 (2015) 103–115.
- [97] S. Zhu, S. Tang, F. Su, Dioscin inhibits ischemic stroke-induced inflammation through inhibition of the TLR4/MyD88/NF-κB signaling pathway in a rat model, Mol. Med. Rep. 17 (2018) 660–666.
- [98] S. Wu, H. Xu, J. Peng, C. Wang, Y. Jin, K. Liu, et al., Potent anti-inflammatory effect of dioscin mediated by suppression of TNF-alpha-induced VCAM-1, ICAM-1 and EL expression via the NF-kappaB pathway, Biochimie 110 (2015) 62–72.
- [99] J. Su, Y. Wei, M. Liu, T. Liu, J. Li, Y. Ji, et al., Anti-hyperuricemic and nephroprotective effects of *Rhizoma Dioscoreae* septemlobae extracts and its main component dioscin via regulation of mOAT1, mURAT1 and mOCT2 in hypertensive mice, Arch. Pharm. Res. 37 (2014) 1336–1344.
- [100] M. Qi, L. Zheng, Y. Qi, X. Han, Y. Xu, L. Xu, et al., Dioscin attenuates renal ischemia/reperfusion injury by inhibiting the TLR4/MyD88 signaling pathway via up-regulation of HSP70, Pharmacol. Res. 100 (2015) 341–352.
- [101] M. Qi, L. Yin, L. Xu, X. Tao, Y. Qi, X. Han, et al., Dioscin alleviates lipopolysaccharide-induced inflammatory kidney injury via the microRNA let-7i/TLR4/ MyD88 signaling pathway, Pharmacol. Res. 111 (2016) 509–522.
- [102] Y. Zhang, X. Tao, L. Yin, L. Xu, Y. Xu, Y. Qi, et al., Protective effects of dioscin against cisplatin-induced nephrotoxicity via the microRNA-34a/sirtuin 1 signalling pathway, Br. J. Pharmacol. 174 (2017) 2512–2527.
- [103] Y. Zhang, Y. Xu, Y. Qi, L. Xu, S. Song, L. Yin, et al., Protective effects of dioscin against doxorubicin-induced nephrotoxicity via adjusting FXR-mediated oxidative stress and inflammation, Toxicology 378 (2017) 53–64.
- [104] Y. Qiao, L. Xu, X. Tao, L. Yin, Y. Qi, Y. Xu, et al., Protective effects of dioscin against fructose- induced renal damage via adjusting Sirt3-mediated oxidative stress, fibrosis, lipid metabolism and inflammation, Toxicol. Lett. 284 (2017) 37–45.

- [105] H.J. Lee, J.S. Park, Y.P. Yoon, Y.J. Shin, S.K. Lee, Y.S. Kim, et al., Dioscin and methylprotodioscin isolated from the root of Asparagus cochinchinensis suppressed the gene expression and production of airway MUC5AC mucin induced by phorbol ester and growth factor, Phytomedicine 22 (2015) 568–572.
- [106] X. Li, G. An, Y. Wang, D. Liang, Z. Zhu, X. Lian, et al., Anti-fibrotic effects of bone morphogenetic protein-7- modified bone marrow mesenchymal stem cells on silica- induced pulmonary fibrosis, Exp. Mol. Pathol. 102 (2017) 70–77.
- [107] C. Li, Y. Lu, S. Du, S. Li, Y. Zhang, F. Liu, et al., Dioscin exerts protective effects against crystalline silica-induced pulmonary fibrosis in mice, Theranostics 7 (2017) 4255–4275.
- [108] H. Zeng, L. Yang, X. Zhang, Y. Chen, J. Cai, Dioscin prevents LPS induced acute lung injury through inhibiting the TLR4/MyD88 signaling pathway via upregulation of HSP70, Mol. Med. Rep. (2018) [Epub ahead of print].
- [109] Y. Hu, X. Tao, X. Han, L. Xu, L. Yin, Y. Qi, et al., Dioscin attenuates gastric ischemia/reperfusion injury through the down-regulation of PKC/ERK1/2 signaling via PKCalpha and PKCbeta2 inhibition, Chem. Biol. Interact. 258 (2016) 234–244.
- [110] O. Escribano, N. Beneit, C. Rubio-Longas, A.R. Lopez-Pastor, A. Gomez-Hernandez, The role of insulin receptor isoforms in diabetes and its metabolic and vascular complications, J. Diabetes Res. 2017 (2017) 1403206.
- [111] H. Sorkhi, N. Saeedizand, M. Poornasrollah, A. Bijani, H. Shafi, Efficacy of potassium polycitrate on renal stone and microlithiasis predisposed by metabolic disorders, Caspian. J. Intern. Med. 8 (2017) 296–300.
- [112] Y.S. Lim, W.R. KiM, The global impact of hepatic fibrosis and end-stage liver disease, Clin. Liver. Dis. 12 (2008) 733–746.
- [113] L. Xu, Y. Wei, D. Dong, L. Yin, Y. Qi, X. Han, et al., iTRAQ-based proteomics for studying the effects of dioscin against nonalcoholic fatty liver disease in rats, RSC Adv. 4 (2014) 30704.
- [114] M. Liu, L. Xu, L. Yin, Y. Qi, Y. Xu, X. Han, et al., Potent effects of dioscin against obesity in mice, Sci. Rep. 5 (2015) 7973.
- [115] B. Poudel, S.W. Lim, H.H. Ki, S. Nepali, Y.M. Lee, D.K. Kim, Dioscin inhibits adipogenesis through the AMPK/MAPK pathway in 3T3-L1 cells and modulates fat accumulation in obese mice, Int. J. Mol. Med. 34 (2014) 1401–1408.
- [116] E.M. Lewiecki, New targets for intervention in the treatment of postmenopa- usal osteoporosis, Nat. Rev. Rheumatol. 7 (2011) 631–638.
- [117] X. Qu, Z. Zhai, X. Liu, H. Li, Z. Ouyang, C. Wu, et al., Dioscin inhibits osteoclast differentiation and bone resorption though down-regulating the akt signaling cascades, Biochem. Biophys. Res. Commun. 443 (2014) 658–665.
- [118] X. Tao, Y. Qi, L. Xu, L. Yin, X. Han, Y. Xu, et al., Dioscin reduces ovariectomyinduced bone loss by enhancing osteoblastogenesis and inhibiting osteoclastogenesis, Pharmacol. Res. 108 (2016) 90–101.
- [119] C. Zhang, J. Peng, S. Wu, Y. Jin, F. Xia, C. Wang, et al., Dioscin promotes osteoblastic proliferation and differentiation via Lrp5 and ER pathway in mouse and human osteoblast-like cell lines, J. Biomed. Sci. 21 (2014) 30.
- [120] C. Zhu, N. Bao, S. Chen, J. Zhao, Dioscin enhances osteoblastic cell differentia- tion and proliferation by inhibiting cell autophagy via the ASPP2/ NF- kappabeta pathway, Mol. Med. Rep. 16 (2017) 4922–4926.
- [121] S. Petta, C. Cammà, D. Cabibi, V. Di Marco, A. Craxì, Hyperuricemia is associated with histological liver damage in patients with non-alcoholic fatty liver disease, Aliment. Pharmacol. Ther. 34 (2011) 757–766.
- [122] S. Yu, H. Yang, X. Guo, X. Zhang, Y. Zhou, Q. Ou, et al., Prevalence of hyperuricemia and its correlates in rural northeast Chinese population: from lifestyle risk factors to metabolic comorbidities, Clin. Rheumatol. 35 (2016) 1207–1215.
- [123] Y. Zhang, L. Jin, J. Liu, W. Wang, H. Yu, J. Li, et al., Effect and mechanism of

dioscin from *Dioscorea spongiosa* on uric acid excretion in animal model of hyperuricemia, J. Ethnopharmacol. 214 (2017) 29–36.

- [124] E. Kurosaki, H. Ogasawara, Ipragliflozin and other sodium-glucose cotranspor- ter-2 (SGLT2) inhibitors in the treatment of type 2 diabetes: preclinical and clinical data, Pharmacol. Ther. 139 (2013) 51–59.
- [125] H. Yu, L. Zheng, L. Xu, L. Yin, Y. Lin, H. Li, et al., Potent effects of the total saponins from *Dioscorea nipponica* Makino against streptozotocin-induced type 2 diabetes mellitus in rats, Phytother. Res. 29 (2015) 228–240.
- [126] H. Jalal, D.F. Bibby, J.W. Tang, J. Bennett, C. Kyriakou, K. Peggs, et al., First reported outbreak of diarrhea due to adenovirus infection in a hematology unit for adults, J. Clin. Microbiol. 43 (2005) 2575–2580.
- [127] G.J. Letchworth, L.L. Rodriguez, J. Del cbarrera, Vesicular stomatitis, Veterinary J. 157 (1999) 239–260.
- [128] M.M. Hassan, D. Li, A.S. El-Deeb, R.A. Wolff, M.L. Bondy, M. Davila, et al., Association between hepatitis B virus and pancreatic cancer, J. Clin. Oncol. 26 (2008) 4557–4562.
- [129] C. Liu, Y. Wang, C. Wu, R. Pei, J. Song, S. Chen, et al., Dioscin's antiviral effect in vitro, Virus Res. 172 (2013) 9–14.
- [130] W.S. Sung, I.S. Lee, D.G. Lee, Damage to the cytoplasmic membrane and cell death caused by lycopene in candida albicans, J. Microbiol. Biotechnol. 17 (2007) 1797–1804.
- [131] J. Cho, H. Choi, J. Lee, M.S. Kim, H.Y. Sohn, D.G. Lee, The antifungal activity and membrane-disruptive action of dioscin extracted from Dioscorea nipponica, Biochimica. et. Biophysica. Acta. 1828 (2013) 1153–1158.
- [132] Y. Guo, E. Xing, H. Song, G. Feng, X. Liang, G. An, et al., Therapeutic effect of dioscin on collagen-induced arthritis through reduction of Th1/Th2, Int. Immunopharmacol. 39 (2016) 79–83.
- [133] G. Feng, Y. Guo, E. Xing, H. Song, A. Gao, X. Zhao, et al., Effects of the dioscin on the expression of ror γ t and Foxp3 in mouse with collagen-induced arthritis, Immunol. J. 31 (2015) 1077–1080.
- [134] C. Chu, F. Shen, H. Zhang, P. Bu, Therapeutic effect of dioscin on collagen- induced arthritis in rats and its mechanisms, J. Clin. Med. 17 (2013) 5–9.
- [135] C. Chu, F. Shen, H. Zhang, P. Bu, Inhibitory effect of dioscin on cyclooxygenase-2 and NF-kB in collagen-induced arthritis rats, Chin. J. Pharmacol. Toxicol. 27 (2013) 341–345.
- [136] H.Y. Lee, D.Y. Jung, H. Ha, K.H. Son, S.J. Jeon, C. Kim, Induction of growth hormone release by dioscin from *Dioscorea batatas* DECNE, J. Biochem. Mol. Biol. 40 (2007) 1016–1020.
- [137] X. Cheng, X. Su, X. Chen, H. Zhao, C. Bo, J. Xu, et al., Biological ingredient analysis of traditional Chinese medicine preparation based on high-throughput sequencing: the story for Liuwei Dihuang Wan, Sci. Rep. 4 (2014) 5147.
- [138] W. Zhou, X. Cheng, Y. Zhang, Effect of Liuwei Dihuang decoction, a traditional Chinese medicinal prescription, on the neuroendocrine immunomodulation network, Pharmacol. Ther. 162 (2016) 170–178.
- [139] Y. Jia, C. Chen, C.S. Ng, S.W. Leung, Meta-analysis of randomized controlled trials on the efficacy of Di'ao Xinxuekang capsule and isosorbide dinitrate in treating angina pectoris, Evid. Based Complement. Alternat Med. 2012 (2012) 904147.
- [140] H. Yu, L. Zheng, L. Yin, L. Xu, Y. Qi, X. Han, et al., Protective effects of the total saponins from *Dioscorea nipponica* Makino against carbon tetrachloride- induced liver injury in mice through suppression of apoptosis and inflammation, Int. Immunopharmacol. 19 (2014) 233–244.
- [141] Z.H. Zhang, Z.J. Chen, Y.C. Sheng, Y.F. Guo, W. Xie, R. Ji, et al., Systematic evaluation of Shuyu Zaogan tablets for coronary heart disease, World Clin. Drugs 32 (2011) 159–164.